



**University of  
Zurich**<sup>UZH</sup>

**Zurich Open Repository and  
Archive**

University of Zurich  
University Library  
Strickhofstrasse 39  
CH-8057 Zurich  
[www.zora.uzh.ch](http://www.zora.uzh.ch)

---

Year: 2015

---

## Hedgehog signaling in basal cell carcinoma

Otsuka, Atsushi ; Levesque, Mitchell P ; Dummer, Reinhard ; Kabashima, Kenji

**Abstract:** Basal cell carcinoma (BCC), the most common type of skin cancer, is occasionally aggressive with deep invasion, destruction of adjacent structures, recurrence and, on very rare occasions, regional and distant metastases. Mutations that occur in BCC in hedgehog (Hh) pathway genes primarily involve the genes encoding patched homolog (PTCH) and smoothened homolog (SMO). Several animal models have demonstrated the functional relevance of genetic alterations in the Hh pathway during tumorigenesis. Recently, targeted therapy has become available both commercially and in the context of human clinical trials. Interestingly, Hh pathway inhibitors not only suppress BCC progression but also promote acquired immune responses. Since immune responses are crucial for long-term tumor control, new clinical trials, such as those involving a combination of Hh inhibitors with immune modifiers, are needed to supplement standard methods of tumor control.

DOI: <https://doi.org/10.1016/j.jdermsci.2015.02.007>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-116903>

Journal Article

Accepted Version



The following work is licensed under a Creative Commons: Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.

Originally published at:

Otsuka, Atsushi; Levesque, Mitchell P; Dummer, Reinhard; Kabashima, Kenji (2015). Hedgehog signaling in basal cell carcinoma. *Journal of Dermatological Science*, 78(2):95-100.

DOI: <https://doi.org/10.1016/j.jdermsci.2015.02.007>

# **Hedgehog signaling in basal cell carcinoma**

Atsushi Otsuka, MD, PhD<sup>1</sup>; Mitchell P. Levesque PhD<sup>2</sup>; Reinhard Dummer, MD<sup>2</sup>; and  
Kenji Kabashima, MD, PhD<sup>1</sup>

<sup>1</sup>Department of Dermatology, Kyoto University Graduate School of Medicine, Kyoto  
606-8507, Japan

<sup>2</sup>Department of Dermatology, University Hospital Zurich, Gloriastrasse 31, 8091 Zurich,  
Switzerland

Correspondence to Atsushi Otsuka, MD, PhD

Department of Dermatology, Kyoto University Graduate School of Medicine

54 Shogoin Kawara, Sakyo-ku, Kyoto 606-8507, Japan

Tel: +81-75-751-3310, Fax: +81-75-761-3002, Email: [otsukamn@kuhp.kyoto-u.ac.jp](mailto:otsukamn@kuhp.kyoto-u.ac.jp)

## **ABSTRACT**

Basal cell carcinoma (BCC), the most common type of skin cancer, is occasionally aggressive with deep invasion, destruction of adjacent structures, recurrence and, on very rare occasions, regional and distant metastases. Mutations that occur in BCC in hedgehog (Hh) pathway genes primarily involve the genes encoding patched homolog (PTCH) and smoothened homolog (SMO). Several animal models have demonstrated the functional relevance of genetic alterations in the Hh pathway during tumorigenesis. Recently, targeted therapy has become available both commercially and in the context of human clinical trials. Interestingly, Hh pathway inhibitors not only suppress BCC progression but also promote acquired immune responses. Since immune responses are crucial for long-term tumor control, new clinical trials, such as those involving a combination of Hh inhibitors with immune modifiers, are needed to supplement standard methods of tumor control.

## 1    **Introduction**

2    Basal cell carcinoma (BCC) is the most common cancer type and its incidence rate is  
3    increasing [1]. BCC characteristically arises in body areas that are exposed to the sun  
4    and is most common on the head and neck (80%), followed by the trunk (15%) and  
5    arms and legs [2]. BCC has also been reported in unusual sites, including the axillae,  
6    breasts, perianal area, genitalia, palms, and soles [3].

7        BCC is generally characterized by slow growth and minimal soft tissue invasiveness  
8    [4]. Since BCC has low metastatic potential, treatment focuses on local  
9    control. Treatment of BCC can be surgical or nonsurgical, such as conventional surgical  
10   excision or micrographic surgery, radiotherapy, photodynamic therapy, cryosurgery, or  
11   topical treatment, including 5-fluorouracil or toll-like receptor agonist imiquimod [5].

12        BCC is occasionally aggressive with deep invasion, destruction of adjacent  
13   structures, recurrence and, on very rare occasions, regional and distant metastasis [6]. A  
14   previous report showed that BCCs comprised 6.6% moderate (640 of 9652) and 0.6%  
15   (58 of 9652) of severe cases [7]. In 2012, the United States Food and Drug  
16   Administration (FDA) approved vismodegib as a first-generation hedgehog (Hh)  
17   pathway antagonist for the treatment of advanced or metastatic BCC. Vismodegib was  
18   also approved in the European Union, Switzerland, Canada, Australia, Mexico, Israel,  
19   South Korea and other countries in 2013. Vismodegib is an effective therapy that  
20   shrinks tumors to a manageable size. In this review, we will discuss the Hh pathway in  
21   BCC and new insights into Hh pathway inhibitors in adaptive immunity for BCC  
22   treatment.

## **Hh signaling**

The Hh pathway plays a crucial role in patterning and organogenesis during early development, and is largely inactive in adults, except for its function in tissue repair and maintenance [8]. The central components of the Hh pathway consist of three secreted ligands (Sonic Hh, Indian Hh, and Desert Hh), a negative regulatory receptor (Patched [PTCH]), a positive regulatory receptor (smoothened [SMO]), and glioma-associated oncogene (GLI) transcription factors (GLI1, GLI2, and GLI3) [8, 9]. The primary cilium is a microtubule-based organelle that protrudes from the plasma membrane and acts as a sensor for extracellular signals, including the Hh pathway [10].

The precise mechanism of Hh signaling through SMO has been well studied. In the absence of Hh ligand, PTCH localizes in the cilia and represses SMO activity by preventing its trafficking and localization to the cilia (Figure 1A). GLI transcription factors are sequestered in the cytoplasm by several protein mediators, including protein kinase A (PKA) and suppressor of fused (SUFU) [11]. GLI undergoes proteasomal cleavage and the resulting repressor from GLI translocates to the nucleus and inhibits the translation of Hh target genes. On ligand binding, PTCH is displaced from the cilia, thereby allowing ciliary accumulation and activation of SMO. Activated GLIs, the final effectors of the pathway, translocate into the nucleus to induce the expression of various context-specific genes, which regulate cellular differentiation, proliferation, and survival (Figure 1B) [11].

## **Hh signaling in BCC**

1 The relationship of Hh pathway activation and cancer has been examined since the  
2 report of germline loss-of-function mutations in PTCH in patients with nevoid basal cell  
3 carcinoma syndrome (NBCCS, Gorlin syndrome) [12]. NBCCS is an  
4 autosomal-dominant disease that is characterized by multiple developmental  
5 abnormalities and a predisposition to tumors, specifically BCC, medulloblastoma (MB),  
6 embryonal rhabdomyosarcoma, and meningioma [13]. Somatic mutations  
7 in PTCH have been identified in 90% of sporadic BCC [14], and gain-of-function  
8 mutations in SMO have been detected in BCC [15]. In particular, recurrent mutations  
9 in SMO and functional studies have demonstrated that these mutations, leading to  
10 aberrant activation of Hh signaling, promote tumor development (Figure 2A) [15].  
11 Recently, Hh pathway mutations have been identified in large-scale whole-genome and  
12 whole-exome deep-sequencing studies across a wide range of cancers. Interestingly,  
13 somatic mutations in PTCH have been detected in other cancer types, such as ovarian  
14 and endometrial cancers [16]. In contrast with BCC and MB, these mutations are  
15 mainly missense. Their relevance in tumor development remains to be determined.

16 In addition to the SMO-dependent pathway, phosphatidylinositol 3-kinase (PI3K)  
17 also promotes Hh signaling in oncogenesis. S6 kinase 1 (S6K1) and atypical protein  
18 kinase C (aPKC), components that are downstream from PI3K, are reported to promote  
19 GLI-dependent transcription. S6K1 is also downstream of the mammalian target of  
20 rapamycin (mTOR) pathway and was found to be elevated in esophageal cancers  
21 resistant to SMO antagonists [17]. In addition, PI3K can promote  
22 3-phosphoinositide-dependent kinase 1 (PDK1) activation and PDK1 can promote

mTOR and S6K1 activation. S6K1 promotes GLI-dependent transcription by phosphorylating GLI1, which prevents an inhibitory interaction with SUFU that allows GLI to enter into the nucleus and turn on target genes. aPKC is an Hh target gene that phosphorylates GLI1 at distinct sites from S6K1, activating GLI1 DNA binding and transcriptional activity to generate a positive-feedback loop that amplifies GLI-dependent transcription in BCC (Figure 2B) [17].

## **Mouse model of BCC**

Several animal models have demonstrated the functional relevance of genetic alterations in the Hh pathway during tumorigenesis. Two spontaneous PTCH mutant animals have been reported, but these mice, such as PTCH<sup>mes/mes</sup> mice, do not develop BCC even after exposure to radiation in spite of these skin anomalies [18, 19]. On the other hand, two different conventional PTCH knockout mouse models, the PTCH<sup>neo12</sup> and PTCH<sup>neo67</sup> strains, in which exons 1 and 2 or exons 6 and 7, respectively, develop BCC [18, 20]. In addition to the several PTCH knockout mice that are known as BCC models, another BCC model exists that includes mice overexpressing Hh, oncogenic SMO, GLI1, or GLI2 specifically in the skin using the keratin (K) 5, 6, or 14 promoters [18]. The skin tumor subtypes range from follicular hamartoma and trichoepithelioma to nodular or invasive BCC depending on the gene and the targeted cell type [18].

## **Hh antagonist**

1 The first well-studied therapy targeting the Hh pathway was cyclopamine, an  
2 endogenous steroidal plant alkaloid derived from corn lilies [21]. Since cyclopia is one  
3 of the defects in mice lacking sonic Hh, this phenotype provided an important  
4 connection between cyclopamine and Hh pathway activation [22]. A number of SMO  
5 antagonists, including vismodegib, were identified using *in vitro* screens to sift through  
6 thousands of compounds (Table 1, Figure 3).

7 Vismodegib is effective at suppressing BCC tumor growth and appears both  
8 tumoricidal and tumoristatic. Most BCC relapses after cessation of vismodegib,  
9 suggesting that the most efficient use of vismodegib as a therapeutic agent is to shrink  
10 tumors to a manageable level and then surgically excise any remaining tumor clones.  
11 Currently, a global single-arm open-label safety study on vismodegib in patients with  
12 advanced BCC (STEVIE study) is ongoing. SMO inhibitors that are in phase I or II  
13 clinical trials to treat advanced or metastatic BCC include sonidegib (LDE225),  
14 erismodegib, XL-139, LEQ506, itraconazole, and saridegib [16, 23]. Furthermore,  
15 there are several candidate components for BCC treatment, such as BEZ235, an  
16 inhibitor of mTOR signaling [24].

17 A previous study demonstrated that vismodegib showed a 30 and 60% response rate  
18 for metastatic and locally advanced BCC, respectively [25]. However, most responses  
19 were only partial. The most sensitive patient population were NBCCS patients who  
20 carry a PTCH mutation that predisposes them to developing hundreds of BCCs.  
21 NBCCS patients treated with vismodegib showed a 100% response rate.  
22 Slower-evolving tumors with low mutation rates, such as sporadic BCC or NBCCS,



1 will respond well to SMO inhibition, whereas metastatic BCCs with higher mutational  
2 rates have a higher likelihood of acquired resistance before or during drug treatment.

3 The most common toxicities reported with Hh pathway inhibitors include taste  
4 alteration (dysgeusia), alopecia, muscle spasms, anorexia, and fatigue [25]. However,  
5 the dose-limiting toxicities are highly variable across the class, probably because of  
6 differences in their structure-activity relationships, on-target potencies, or tissue  
7 distributions.

#### 8 **Promotion of acquired immune response by Hh signal inhibitors**

10 Cancers have several mechanisms to escape immune surveillance [26]. Despite the  
11 presence of cancer-testis and other tumor antigens, BCC also escapes immune  
12 surveillance through the down-regulation of HLA class I expression [27]. Recently, we  
13 demonstrated that vismodegib promotes acquired immune response. [In the context of](#)  
14 HLA class I expression, cytotoxic CD8<sup>+</sup> T cells play essential roles in anti-tumor effects  
15 for skin tumors [28, 29]. We have shown substantial alterations in the immune-  
16 microenvironment with an intra- and peritumoral increase of cytotoxic CD8<sup>+</sup> T cells and  
17 an up-regulation of MHC class I during tumor regression under treatment with Hh  
18 pathway inhibitors [30]. Moreover, a reduction in primary cilia was observed after Hh  
19 pathway inhibitor treatment [30]. Before treatment, all BCC cells are ciliated,  
20 suggesting that they are responsive to Hh signaling. By inhibiting Hh signaling, the  
21 BCC cells lose their cilia and subsequently stop proliferating.

22 T cell activation requires both T cell receptor (TCR) and co-stimulatory molecule

1 ligation by professional antigen-presenting cells (APC), and the outcome of the  
2 stimulatory signal is influenced by the microenvironment of the T cell and the APC [31,  
3 32]. The Hh signaling pathway reduced the strength of the TCR signal in mature  
4 peripheral T cells [33, 34]. Additionally, the repression of the Hh signaling pathway in  
5 T cells increased T cell activation [35]. This suggests that the Hh pathway inhibitor has  
6 direct effects on peripheral T cell and activates adaptive immune responses.

7 It is known that IFN- $\gamma$  up-regulates MHC class I antigen presentation by inducing  
8 gene expression signatures that are related to MHC class I antigen processing and  
9 presentation, including activation of the JAK/STAT1 signal transduction pathway  
10 (Figure 4A) [36]. Potential cross-talk between IFN- $\gamma$  and the Hh pathway was recently  
11 described by Laner-Plamberger et al. [37]. They demonstrated that the suppressor of  
12 cytokine signaling 1 (SOCS1) is a direct target of Hh/GLI signaling in human  
13 keratinocytes and medulloblastoma cells and a potent inhibitor of IFN- $\gamma$ -STAT1  
14 signaling, which can induce cell cycle arrest, apoptosis, and anti-tumor immunity. It was  
15 shown that the transcription factors GLI1 and GLI2 activated the SOCS1 promoter and  
16 that STAT1 phosphorylation was reduced in cells with active Hh/GLI signaling (Figure  
17 4B) [37]. During treatment with Hh pathway inhibitors, GLI was suppressed by  
18 proteosomal cleavage and did not activate the SOCS1 promoter. Up-regulation of MHC  
19 class I after treatment with Hh pathway inhibitors may be induced by this mechanism. To  
20 understand the precise mechanism, further studies are needed in the future.

## 21 22 **Conclusion**

1 BCCs are highly prevalent tumors that are treatable using traditional therapy, including  
2 both surgical and nonsurgical methods. Traditional therapies are not as effective,  
3 however, in treating multiple BCCs or those that become highly invasive or  
4 metastatic. The advancement of Hh pathway inhibitors, such as vismodegib, into  
5 clinical development has yielded good responses in mutation-driven tumors with  
6 activated Hh signaling. However, it appears that the clinical application of single-agent  
7 Hh pathway inhibitors might not be as broad as was initially expected.

8 In addition to the effect of Hh pathway inhibitors on BCC proliferation, we  
9 demonstrated that Hh pathway inhibitor treatment induced a recruitment of cytotoxic T  
10 cells into the tumor and up-regulation of MHC class I in BCCs. Reduction in the  
11 frequency of ciliated cells during the Hh pathway inhibitor treatment suggests that cilia  
12 are required for Hh inhibitor efficacy. For long-term tumor control, immune responses  
13 are crucial. It has been reported that immune modifiers, including imiquimod, are  
14 therapeutically beneficial [38]. New clinical trials, such as those involving a  
15 combination of Hh inhibitors with immune modifiers, are needed to supplement  
16 standard methods of tumor control.

## 19 **ACKNOWLEDGEMENTS**

20 This work was supported in part by Grants-in-Aid for Scientific Research from the  
21 Ministry of Education, Culture, Sports, Science and Technology and the Ministry of  
22 Health, Labour and Welfare of Japan. No additional external funding was received for

this study.

## References

- [1] Lomas A, Leonardi-Bee J, Bath-Hextall F: A systematic review of worldwide incidence of nonmelanoma skin cancer. *Br J Dermatol* 166: 1069-1080, 2012.
- [2] Rubin AI, Chen EH, Ratner D: Basal-cell carcinoma. *N Engl J Med* 353: 2262-2269, 2005.
- [3] Dreier J, Cheng PF, Bogdan Alleman I, Gugger A, Hafner J, Tschopp A, et al.: Basal cell carcinomas in a tertiary referral centre- a systematic analysis. *Br J Dermatol*, 2014.
- [4] Walling HW, Fosko SW, Geraminejad PA, Whitaker DC, Arpey CJ: Aggressive basal cell carcinoma: presentation, pathogenesis, and management. *Cancer Metastasis Rev* 23: 389-402, 2004.
- [5] Arits AH, Mosterd K, Essers BA, Spoorenberg E, Sommer A, De Rooij MJ, et al.: Photodynamic therapy versus topical imiquimod versus topical fluorouracil for treatment of superficial basal-cell carcinoma: a single blind, non-inferiority, randomised controlled trial. *Lancet Oncol* 14: 647-654, 2013.
- [6] Boaventura P, Pereira D, Mendes A, Batista R, da Silva AF, Guimaraes I, et al.: Mitochondrial D310 D-Loop instability and histological subtypes in radiation-induced cutaneous basal cell carcinomas. *J Dermatol Sci* 73: 31-39, 2014.
- [7] Dreier J, Cheng PF, Bogdan Alleman I, Gugger A, Hafner J, Tschopp A, et al.: Basal cell carcinomas in a tertiary referral centre: a systematic analysis. *Br J Dermatol* 171: 1066-1072, 2014.
- [8] Varjosalo M, Taipale J: Hedgehog: functions and mechanisms. *Genes Dev* 22: 2454-2472, 2008.
- [9] Johnson RL, Rothman AL, Xie J, Goodrich LV, Bare JW, Bonifas JM, et al.: Human homolog of patched, a candidate gene for the basal cell nevus syndrome. *Science* 272: 1668-1671, 1996.
- [10] Wong SY, Reiter JF: The primary cilium at the crossroads of mammalian hedgehog signaling. *Curr Top Dev Biol* 85: 225-260, 2008.
- [11] Amakye D, Jagani Z, Dorsch M: Unraveling the therapeutic potential of the Hedgehog pathway in cancer. *Nat Med* 19: 1410-1422, 2013.

- [12] Hahn H, Wicking C, Zaphiropoulous PG, Gailani MR, Shanley S, Chidambaram A, et al.: Mutations of the human homolog of *Drosophila* patched in the nevoid basal cell carcinoma syndrome. *Cell* 85: 841-851, 1996.
- [13] Gorlin RJ: Nevoid basal cell carcinoma syndrome. *Dermatol Clin* 13: 113-125, 1995.
- [14] Gailani MR, Stahle-Backdahl M, Leffell DJ, Glynn M, Zaphiropoulos PG, Pressman C, et al.: The role of the human homologue of *Drosophila* patched in sporadic basal cell carcinomas. *Nat Genet* 14: 78-81, 1996.
- [15] Xie J, Murone M, Luoh SM, Ryan A, Gu Q, Zhang C, et al.: Activating Smoothed mutations in sporadic basal-cell carcinoma. *Nature* 391: 90-92, 1998.
- [16] Atwood SX, Whitson RJ, Oro AE: Advanced treatment for basal cell carcinomas. *Cold Spring Harb Perspect Med* 4: a013581, 2014.
- [17] Athar M, Li C, Kim AL, Spiegelman VS, Bickers DR: Sonic hedgehog signaling in Basal cell nevus syndrome. *Cancer Res* 74: 4967-4975, 2014.
- [18] Nitzki F, Becker M, Frommhold A, Schulz-Schaeffer W, Hahn H: Patched knockout mouse models of Basal cell carcinoma. *J Skin Cancer* 2012: 907543, 2012.
- [19] Makino S, Masuya H, Ishijima J, Yada Y, Shiroishi T: A spontaneous mouse mutation, mesenchymal dysplasia (mes), is caused by a deletion of the most C-terminal cytoplasmic domain of patched (ptc). *Dev Biol* 239: 95-106, 2001.
- [20] Hahn H, Wojnowski L, Zimmer AM, Hall J, Miller G, Zimmer A: Rhabdomyosarcomas and radiation hypersensitivity in a mouse model of Gorlin syndrome. *Nat Med* 4: 619-622, 1998.
- [21] Keeler RF, Binns W: Teratogenic compounds of *Veratrum californicum* (Durand). I. Preparation and characterization of fractions and alkaloids for biologic testing. *Can J Biochem* 44: 819-828, 1966.
- [22] Chiang C, Litingtung Y, Lee E, Young KE, Corden JL, Westphal H, et al.: Cyclopia and defective axial patterning in mice lacking Sonic hedgehog gene function. *Nature* 383: 407-413, 1996.
- [23] Dreier J, Dummer R, Felderer L, Nageli M, Gobbi S, Kunstfeld R: Emerging drugs and combination strategies for basal cell carcinoma. *Expert Opin Emerg Drugs* 19: 353-365, 2014.
- [24] Liang M, Lv J, Chu H, Wang J, Chen X, Zhu X, et al.: Vertical inhibition of PI3K/Akt/mTOR signaling demonstrates in vitro and in vivo anti-fibrotic activity. *J*

1 Dermatol Sci 76: 104-111, 2014.

2 [25] Sekulic A, Migden MR, Oro AE, Dirix L, Lewis KD, Hainsworth JD, et al.:  
3 Efficacy and safety of vismodegib in advanced basal-cell carcinoma. N Engl J Med 366:  
4 2171-2179, 2012.

5 [26] Gehrke S, Otsuka A, Huber R, Meier B, Kistowska M, Fenini G, et al.: Metastatic  
6 melanoma cell lines do not secrete IL-1beta but promote IL-1beta production from  
7 macrophages. J Dermatol Sci 74: 167-169, 2014.

8 [27] Walter A, Barysch MJ, Behnke S, Dziunycz P, Schmid B, Ritter E, et al.:  
9 Cancer-testis antigens and immunosurveillance in human cutaneous squamous cell and  
10 basal cell carcinomas. Clin Cancer Res 16: 3562-3570, 2010.

11 [28] Fujiyama T, Oze I, Yagi H, Hashizume H, Matsuo K, Hino R, et al.: Induction of  
12 cytotoxic T cells as a novel independent survival factor in malignant melanoma with  
13 percutaneous peptide immunization. J Dermatol Sci 75: 43-48, 2014.

14 [29] Kato Y, Yoshino I, Egusa C, Maeda T, Pili R, Tsuboi R: Combination of HDAC  
15 inhibitor MS-275 and IL-2 increased anti-tumor effect in a melanoma model via  
16 activated cytotoxic T cells. J Dermatol Sci 75: 140-147, 2014.

17 [30] Otsuka A, Dreier J, Cheng PF, Nageli M, Lehmann H, Felderer L, et al.: Hedgehog  
18 pathway inhibitors promote adaptive immune responses in Basal Cell Carcinoma. Clin  
19 Cancer Res, (in press).

20 [31] Nomura T, Kabashima K, Miyachi Y: The panoply of alphabetaT cells in the skin. J  
21 Dermatol Sci 76: 3-9, 2014.

22 [32] Haniffa M, Gunawan M, Jardine L: Human skin dendritic cells in health and  
23 disease. J Dermatol Sci, 2014.

24 [33] Rowbotham NJ, Hager-Theodorides AL, Cebecauer M, Shah DK, Drakopoulou E,  
25 Dyson J, et al.: Activation of the Hedgehog signaling pathway in T-lineage cells inhibits  
26 TCR repertoire selection in the thymus and peripheral T-cell activation. Blood 109:  
27 3757-3766, 2007.

28 [34] Rowbotham NJ, Hager-Theodorides AL, Furmanski AL, Crompton T: A novel role  
29 for Hedgehog in T-cell receptor signaling: implications for development and immunity.  
30 Cell Cycle 6: 2138-2142, 2007.

31 [35] Rowbotham NJ, Furmanski AL, Hager-Theodorides AL, Ross SE, Drakopoulou E,  
32 Koufaris C, et al.: Repression of hedgehog signal transduction in T-lineage cells  
33 increases TCR-induced activation and proliferation. Cell Cycle 7: 904-908, 2008.

- [36] Zhou F: Molecular Mechanisms of IFN- $\gamma$  to Up-Regulate MHC Class I Antigen Processing and Presentation. *International Reviews of Immunology* 28: 239-260, 2009.
- [37] Laner-Plamberger S, Wolff F, Kaser-Eichberger A, Swierczynski S, Hauser-Kronberger C, Frischauf AM, et al.: Hedgehog/GLI signaling activates suppressor of cytokine signaling 1 (SOCS1) in epidermal and neural tumor cells. *PLoS One* 8: e75317, 2013.
- [38] Ogawa Y, Kawamura T, Matsuzawa T, Aoki R, Shimada S: Recruitment of plasmacytoid dendritic cells to skin regulates treatment responsiveness of actinic keratosis to imiquimod. *J Dermatol Sci* 76: 67-69, 2014.
- [39] Rodon J, Tawbi HA, Thomas AL, Stoller RG, Turtzsch CP, Baselga J, et al.: A phase I, multicenter, open-label, first-in-human, dose-escalation study of the oral smoothened inhibitor Sonidegib (LDE225) in patients with advanced solid tumors. *Clin Cancer Res* 20: 1900-1909, 2014.
- [40] Munchhof MJ, Li Q, Shavnya A, Borzillo GV, Boyden TL, Jones CS, et al.: Discovery of PF-04449913, a Potent and Orally Bioavailable Inhibitor of Smoothened. *ACS Med Chem Lett* 3: 106-111, 2012.
- [41] Skvara H, Kalthoff F, Meingassner JG, Wolff-Winiski B, Aschauer H, Kelleher JF, et al.: Topical treatment of Basal cell carcinomas in nevoid Basal cell carcinoma syndrome with a smoothened inhibitor. *J Invest Dermatol* 131: 1735-1744, 2011.
- [42] Lappano R, Maggiolini M: G protein-coupled receptors: novel targets for drug discovery in cancer. *Nat Rev Drug Discov* 10: 47-60, 2011.
- [43] Tremblay MR, Lescarbeau A, Grogan MJ, Tan E, Lin G, Austad BC, et al.: Discovery of a potent and orally active hedgehog pathway antagonist (IPI-926). *J Med Chem* 52: 4400-4418, 2009.
- [44] Kim J, Aftab BT, Tang JY, Kim D, Lee AH, Rezaee M, et al.: Itraconazole and arsenic trioxide inhibit Hedgehog pathway activation and tumor growth associated with acquired resistance to smoothened antagonists. *Cancer Cell* 23: 23-34, 2013.
- [45] Campbell SB, Walker R, Tai SS, Jiang Q, Russ GR: Randomized controlled trial of sirolimus for renal transplant recipients at high risk for nonmelanoma skin cancer. *Am J Transplant* 12: 1146-1156, 2012.

Table 1

Hh pathway antagonists that are currently in use or development for the treatment of BCCs

Inhibitor	Target	Reference
Vismodegib	SMO	[25]
Sonidegib	SMO	[39]
PF-04449913	SMO	[40]
Erismodegib	SMO	[41]
LEQ506	SMO	[42]
Saridegib	SMO	[43]
Itraconazole	SMO	[44]
ATO	GLI	[44]
Sirolimus	mTOR	[45]

SMO, Smoothed; ATO, arsenic trioxide; mTOR, mammalian target of rapamycin

## Figure legends

### Figure 1. Mechanism of the Hh pathway

(A) In the absence of Hh ligand, PTCH localizes in the cilia and represses SMO activity by preventing its trafficking and localization to the cilia. GLI transcription factors are sequestered in the cytoplasm by several protein mediators, including PKA and SUFU.



1 GLI undergoes proteasomal cleavage and the resulting repressor form GLI translocates  
2 to the nucleus and inhibits the translation of Hh target genes. (B) On ligand binding,  
3 PTCH is displaced from the cilia, thereby allowing ciliary accumulation and activation  
4 of SMO. Activated GLIs, the final effectors of the pathway, translocate into the nucleus  
5 to induce the expression of various context-specific genes that regulate cellular  
6 differentiation, proliferation, and survival.

7  
8 **Figure 2. Hh pathway in BCC**

9 (A) Inactivating mutations in PTCH or the binding of Hh ligands to PTCH de-represses  
10 SMO, thereby allowing its translocation onto the tip of the primary cilium, leading to  
11 the transcriptional activation of Gli. Multiple ciliary proteins are involved in processing  
12 Hh signal transduction. The activation and nuclear translocation of Gli involve the  
13 dissociation of Gli from its endogenous inhibitor SUFU. (B) S6K1 and aPKC, which  
14 are downstream of PI3K, are reported to promote GLI-dependent transcription. S6K1 is  
15 also downstream of the mTOR pathway. PI3K can promote PDK1 activation and PDK1  
16 can promote mTOR and S6K1 activation. S6K1 promotes GLI-dependent transcription  
17 by phosphorylating GLI1, which prevents an inhibitory interaction with SUFU that  
18 allows GLI to enter into the nucleus and turn on target genes. aPKC is an Hh target gene  
19 that phosphorylates GLI1 at distinct sites from S6K1, activating GLI1 DNA binding and  
20 transcriptional activity to generate a positive-feedback loop that amplifies  
21 GLI-dependent transcription in BCC.

**Figure 3. Hh pathway antagonists for BCC treatment**

Hh pathway activity can be inhibited through several mechanisms, including direct binding to SMO, modulation of primary cilia translocation of SMO, inhibition of the receptor–ligand interaction, and inhibition of GLI transcription factors.

**Figure 4. Hypothesis of up-regulation of MHC class I during treatment with Hh pathway inhibitors**

(A) IFN- $\gamma$  up-regulates MHC class I antigen presentation by inducing gene expression signatures that are related to MHC class I antigen processing and presentation, including activation of the JAK/STAT1 signal transduction pathway. SOCS1 is a direct target of Hh/GLI signaling in human keratinocytes and medulloblastoma cells and a potent inhibitor of IFN- $\gamma$ -STAT1 signaling, which can induce cell cycle arrest, apoptosis, and anti-tumor immunity. It was shown that the transcription factors GLI1 and GLI2 activated the SOCS1 promoter and that STAT1 phosphorylation was reduced in cells with active Hh/GLI signaling. (B) During treatment with Hh pathway inhibitors, GLI undergoes proteosomal cleavage and does not activate the SOCS1 promoter. Up-regulation of MHC class I after treatment with Hh pathway inhibitors may be induced by this mechanism.